



TITLE:

# <Original>De Novo Synthesis of Veratryl Alcohol by *Coriolus versicolor*

AUTHOR(S):

KAWAI, Shingo; UMEZAWA, Toshiaki; HIGUCHI, Takayoshi

---

CITATION:

KAWAI, Shingo ...[et al]. <Original>De Novo Synthesis of Veratryl Alcohol by *Coriolus versicolor*. Wood research : bulletin of the Wood Research Institute Kyoto University 1986, 73: 18-21

ISSUE DATE:

1986-12-28

URL:

<http://hdl.handle.net/2433/53303>

RIGHT:

## ***De Novo* Synthesis of Veratryl Alcohol by *Coriolus versicolor*\***

Shingo KAWAI\*\*, Toshiaki UMEZAWA\*\*,  
and Takayoshi HIGUCHI\*\*

(Received September 1, 1986)

**Abstract**—Veratryl alcohol was found in ligninolytic culture of *Coriolus versicolor*. The structure of veratryl alcohol synthesized *de novo* was confirmed in comparison with <sup>1</sup>H-NMR spectra of the authentic dimethoxybenzyl alcohols (veratryl alcohol and its isomers).

### **1. Introduction**

Lignin biodegradation has been greatly elucidated in recent years<sup>1-5</sup>). Ligninolytic enzyme (lignin peroxidase, ligninase) was purified from the culture filtrate of *Phanerochaete chrysosporium*<sup>6-9</sup>) and the reaction mechanism of lignin peroxidase *via* aryl cation radical was proposed<sup>10,11</sup>). This enzyme activity was enhanced by the addition of veratryl alcohol, a secondary metabolite of *P. chrysosporium*<sup>12</sup>).

We previously reported the degradation of non-phenolic  $\beta$ -O-4 lignin substructure model compounds in ligninolytic culture of *Coriolus versicolor* and suggested that a similar lignin peroxidase is excreted by *C. versicolor*<sup>13-15</sup>). In the present paper we report *de novo* synthesis of veratryl alcohol by *C. versicolor* and discuss the role of veratryl alcohol in lignin biodegradation.

### **2. Materials and Methods**

#### **2.1 Culture Conditions and Extraction**

*Coriolus versicolor* Ps4a was maintained on 2% malt agar slants. Experimental culture (20 ml in 300 ml-Erlenmeyer flasks) were inoculated with a small mycelial mat from the slant and grown without agitation at 30°C. The culture medium was prepared as described previously<sup>13</sup>).

The 7-day-old cultures (28 cultures) were flushed with sterile oxygen and incubated under the same conditions for 3 days. The whole cultures were combined, acidified with 1N HCl to pH 2 and extracted with 1 liter of ethyl acetate. The

\* A part of this paper was presented at the 29th symposium on lignin in Tokyo, Oct. 1984.

\*\* Research Section of Lignin Chemistry.

organic layer was washed with saturated NaCl solution, dried over anhydrous  $\text{Na}_2\text{SO}_4$  and concentrated to dryness.

## 2.2 Syntheses of Authentic Compounds

2,3-Dimethoxybenzyl alcohol was prepared from 2-hydroxy-3-methoxybenzaldehyde (*o*-vanillin, Nakarai Chemicals Ltd.) *via* the following two steps; (i) methyl iodide/ $\text{K}_2\text{CO}_3$  in DMF at room temperature, and (ii)  $\text{NaBH}_4$  in methanol at  $0^\circ\text{C}$ .

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ )  $\delta$  (ppm); 3.87 (3H, s,  $-\text{OCH}_3$ ), 3.89 (3H, s,  $-\text{OCH}_3$ ), 4.70 (2H, s,  $-\text{CH}_2-$ ), 6.82–7.12 (3H, m, aromatic-H).

2,4-Dimethoxybenzyl alcohol was prepared from 2,4-dihydroxybenzaldehyde ( $\beta$ -resorcyraldehyde, Nakarai Chemicals Ltd.) *via* the following two steps; (i) methyl iodide/ $\text{K}_2\text{CO}_3$  in DMF at room temperature, and (ii)  $\text{NaBH}_4$  in methanol at  $0^\circ\text{C}$ .

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ )  $\delta$  (ppm); 3.80 (3H, s,  $-\text{OCH}_3$ ), 3.83 (3H, s,  $-\text{OCH}_3$ ), 4.60 (2H, s,  $-\text{CH}_2-$ ), 6.40–6.46 (2H, m, aromatic- $\text{H}_{3,5}$ ), 7.16 (1H, d,  $J=9.0$ , aromatic- $\text{H}_6$ ).

2,5-Dimethoxybenzyl alcohol was prepared from 2,5-dimethoxybenzaldehyde (Nakarai Chemicals Ltd.) by reduction with  $\text{NaBH}_4$  in methanol at  $0^\circ\text{C}$ .

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ )  $\delta$  (ppm); 3.77 (3H, s,  $-\text{OCH}_3$ ), 3.81 (3H, s,  $-\text{OCH}_3$ ), 4.65 (2H, s,  $-\text{CH}_2-$ ), 6.76–6.90 (3H, m, aromatic-H).

3,4-Dimethoxybenzyl alcohol (veratryl alcohol) was commercially available (Tokyo Chemical Industry Co., Ltd.).

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ )  $\delta$  (ppm); 3.87 (3H, s,  $-\text{OCH}_3$ ), 3.88 (3H, s,  $-\text{OCH}_3$ ), 4.61 (2H, s,  $-\text{CH}_2-$ ), 6.78–6.94 (3H, m, aromatic-H).

## 2.3 Instrument

$^1\text{H}$ -NMR spectra were obtained with a Varian XL-200 FT-NMR spectrometer (200 MHz) using tetramethylsilane as an internal standard. Chemical shifts and coupling constants are given in  $\delta$  values (ppm) and Hz, respectively.

## 3. Results and Discussion

The extracts were submitted to TLC (Kiesel gel 60,  $\text{F}_{254}$ , Merck, developing solvent:  $\text{CH}_2\text{Cl}_2$ ). Veratryl alcohol was isolated and its structure was identified by  $^1\text{H}$ -NMR. The  $^1\text{H}$ -NMR spectra of the metabolic veratryl alcohol and authentic compounds are shown in Fig. 1. Possibility of 3,5-dimethoxybenzyl alcohol is ruled out, because the protons of the two methoxyl groups of the metabolite have different chemical shifts in  $^1\text{H}$ -NMR spectrum, while the chemical shifts of methoxyl groups of 3,5-dimethoxybenzyl alcohol are identical. From the  $^1\text{H}$ -NMR spectra shown in Fig. 1, it is clear that the metabolic product (A) is veratryl alcohol (B) and not other isomers (C–E).

Russell *et al.*<sup>16)</sup> found veratraldehyde in a culture of *C. versicolor*. *De novo* syn-

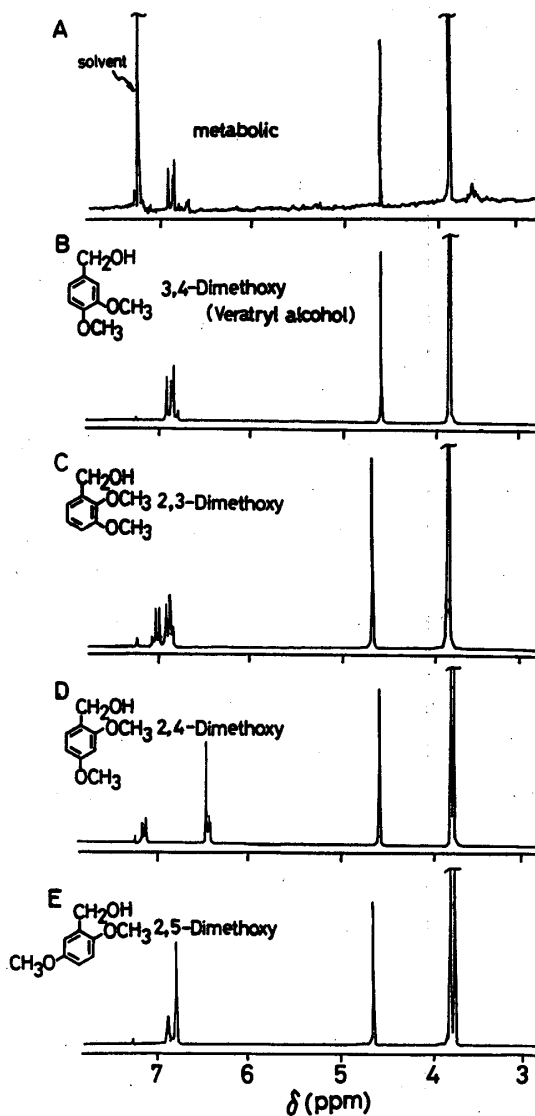


Fig. 1.  $^1\text{H}$ -NMR spectra of metabolic veratryl alcohol (A) and authentic compounds (B-E).

thesis of veratryl alcohol by *P. chrysosporium* was reported previously by Lundquist and Kirk<sup>17)</sup>. Afterwards many papers were published in relation to physiological and biochemical role of veratryl alcohol in lignin biodegradation by *P. chrysosporium*. Shimada *et al.*<sup>18)</sup> reported the biosynthesis of veratryl alcohol in relation to lignin degradation by *P. chrysosporium*. It was demonstrated that addition of veratryl alcohol to the culture of *P. chrysosporium* increased the ligninolytic activity and the production of lignin peroxidase in the culture<sup>12,19)</sup>. It was further reported that the oxidation of monomethoxylated aromatic monomers<sup>20)</sup> and 2-keto-4-thiomethyl butyric acid (KTBA)<sup>21)</sup> by lignin peroxidase of *P. chrysosporium* was enhanced by veratryl alcohol.

Our previous investigations<sup>13~15)</sup> showed that the main degradation products of non-phenolic  $\beta$ -O-4 lignin substructure model compounds by *C. versicolor* were similar to those obtained by lignin peroxidase of *P. chrysosporium*<sup>6~9,22,23)</sup>, suggesting that a similar lignin peroxidase is excreted by *C. versicolor*. These results also suggest that veratryl alcohol enhances the ligninolytic activity and the production of the lignin peroxidase by *C. versicolor*.

### Acknowledgement

This research was partly supported by a Grant-in-Aid for Scientific research (No. 59760124) from the Ministry of Education of Japan.

### References

- 1) C.-L. CHEN and H.-m. CHANG: "Biosynthesis and Biodegradation of Wood Components", (T. HIGUCHI ed) Academic Press, Florida, p.535 (1985).
- 2) T. HIGUCHI: *ibid.* p. 557.
- 3) T.K. KIRK and M. SHIMADA: *ibid.* p. 579.
- 4) M.S.A. LEISOLA and A. FIECHTER: "Advances in Biotechnological Processes vol. 5", (A. MIZRAHI and A.L. VAN WEZAL eds.) Alan R. Liss, New York, p. 59 (1985).
- 5) P.J. HARVEY, H.E. SHOEMAKER and J.M. PALMER: *Ann. Proc. Phytochem. Soc. Eur.*, **26**, 249 (1985).
- 6) M. TIEN and T.K. KIRK: *Science*, **221**, 661 (1983).
- 7) M. TIEN and T.K. KIRK: *Proc. Natl. Acad. Sci. USA*, **81**, 2280 (1984).
- 8) J.K. GLENN, M.A. MORGAN, M.B. MAYFIELD, M. KUWAHARA and M.H. GOLD: *Biochem. Biophys. Res. Commun.*, **114**, 1077 (1983).
- 9) M.H. GOLD, M. KUWAHARA, A.A. CHIU and J.K. GLENN: *Arch. Biochem. Biophys.*, **234**, 353 (1984).
- 10) P.J. KERSTEN, M. TIEN, B. KALYANARAMAN and T.K. KIRK: *J. Biol. Chem.*, **260**, 2609 (1985).
- 11) K.E. HAMMEL, M. TIEN, B. KALYANARAMAN and T.K. KIRK: *J. Biol. Chem.*, **260**, 8348 (1985).
- 12) B.D. FAISON and T.K. KIRK: *Appl. Environ. Microbiol.*, **49**, 299 (1985).
- 13) S. KAWAI, T. UMEZAWA and T. HIGUCHI: *Agric. Biol. Chem.*, **49**, 2325 (1985).
- 14) S. KAWAI, T. UMEZAWA and T. HIGUCHI: *Appl. Environ. Microbiol.*, **50**, 1505 (1985).
- 15) S. KAWAI, T. UMEZAWA and T. HIGUCHI: *FEBS Lett.*, in press.
- 16) J.D. RUSSELL, M.E.K. HENDERSON and V.C. FARMER: *Biochim. Biophys. Acta*, **52**, 565 (1961).
- 17) K. LUNDQUIST and T.K. KIRK: *Phytochem.*, **17**, 1676 (1978).
- 18) M. SHIMADA, F. NAKATSUBO, T.K. KIRK and T. HIGUCHI: *Arch. Microbiol.*, **129**, 321 (1981).
- 19) M.S.A. LEISOLA, D.C. ULMER, R. WALDNER and A. FIECHTER: *J. Biotechnol.*, **1**, 331 (1984).
- 20) P.J. HARVEY, H.E. SHOEMAKER and J.M. PALMER: *FEBS Lett.*, **195**, 242 (1986).
- 21) V. RANGANATHAN, K. MIKI and M.H. GOLD: *Arch. Biochem. Biophys.*, **241**, 304 (1985).
- 22) T. UMEZAWA, M. SHIMADA, T. HIGUCHI and K. KUSAI: *FEBS Lett.*, **205**, 287 (1986).
- 23) T. UMEZAWA and T. HIGUCHI: *FEBS Lett.*, **205**, 293 (1986).